# ASTM E 2149-20

Standard Test Method for Determining Antimicrobial Activity of Antimicrobial Agents Under Dynamic Contact Conditions

FINAL REPORT: R2022-268-4A
\*AMENDMENT TO R2022-268-4

Prepared for:
James Hardie Philippines Inc.
Brgy. San Isidro
Cabuyao, Laguna, Philippines 4025

Accredited Testing Provided by:



130 Erick Street Crystal Lake, IL 60014 815.526.0954 TESTING CERT: #2832.01

Testing Initiated: June 21, 2022 Testing Completed: June 27, 2022 Report Issued: July 20, 2022

Performed By: Agata Shulfer Approved By: Debbie Koester Title: Senior Staff Scientist Title: Quality Manager



# **Objective:**

To determine the antimicrobial activity of antimicrobial agents under dynamic contact conditions of four samples as seen by the ASTM E 2149-20 test method.

# **Test Sample Identification\*:**

- 1. HardieFlex® sample 1
- 2. HardieFlex® sample 2
- 3. HardieFlex® NexGen™ with MoldBlock™ Technology sample 142037
- 4. HardieFlex® NexGen™ with MoldBlock™ Technology sample 142038

# **Test Procedure Summary:**

The test organism was adjusted and diluted to obtain a working bacterial inoculum solution. A flask for each sample and the "inoculum only" was prepared containing 50 mL of the working bacterial inoculum. Serial dilutions of the "0" contact time were performed on the "inoculum only" flask. The flask containing the test sample and the "inoculum only" flask were placed on the wrist action shaker for appropriate contact time. After shaking, serial dilutions were made and the plates incubated. After incubation, colonies of recovered bacteria are counted and used to determine percent and log reductions.

#### Test Variables

| Test Organism:                         | Stachybotrys chartarum ATCC#16026              |  |  |
|--|--|--|--|
| Sample Description:                    | Solid  |  |  |
| Sample Weight/Size:                    | 2" x 2" square cut into smaller pieces         |  |  |
| Method of Sterilization /Pre-Cleaning: | None   |  |  |
| <b>Dilution Medium Used:</b>           | Sterile buffer solution per standard           |  |  |
| Buffer/Shake Solution Used:            | Sterile buffer solution per standard           |  |  |
| Serial Dilution Medium Used:           | D/E Neutralizing Broth                         |  |  |
| Untreated Control:                     | Inoculum Only                                  |  |  |
| Working Inoculum Concentration:        | S. chartarum ATCC#16026: 1.2 x 10 <sup>5</sup> |  |  |
| <b>Contact Time:</b>                   | 24 Hours                                       |  |  |
| Deviations from                        | None, testing performed per ASTM E2149-20      |  |  |
| Standard Test Method:                  | without deviation.                             |  |  |



#### **Test Results:**

The results for the test pieces can be found in the data table below. These results pertain only to the samples tested.

# Results against Stachybotrys chartarum ATCC#16026:

#### Percent reduction of bacteria per sample against inoculum only

| Time = 24 Hours  | Average Number of Bacteria<br>Recovered (spores/ml) | Percent<br>Reduction | Log<br>Reduction |
|--|---|----------------------|------------------|
| Inoculum Only  | $6.6 \times 10^4$                                   |                      |                  |
| HardieFlex® sample 1   | $4.9 \times 10^4$                                   | 26                   | 0.13             |
| HardieFlex® sample 2   | 3.5 x 10 <sup>4</sup>                               | 47                   | 0.28             |
| HardieFlex® NexGen™ with<br>MoldBlock™ Technology<br>sample 142037 | $7.3 \times 10^3$                                   | 89                   | 0.95             |
| HardieFlex® NexGen™ with<br>MoldBlock™ Technology<br>sample 142038 | $5.8 \times 10^{2}$                                 | 99.1                 | 2.06             |

Percent reductions and log reductions are determined by comparing the sample after the contact time to the "inoculum only" after the contact. Percent reduction is translated into log reduction by the following:

90% reduction = 1 log reduction; i.e. 1,000,000 reduced to 100,000 is a 1 log reduction 99% reduction = 2 log reduction; i.e. 1,000,000 reduced to 10,000 is a 2 log reduction 99.9% reduction = 3 log reduction; i.e. 1,000,000 reduced to 1,000 is a 3 log reduction 99.99% reduction = 4 log reduction; i.e. 1,000,000 reduced to 100 is a 4 log reduction 99.99% reduction = 5 log reduction; i.e. 1,000,000 reduced to 10 is a 5 log reduction